

# UbiQ

targeting the ubiquitin system

[www.ubiqbio.com](http://www.ubiqbio.com)

## HTRF K48 linked diUbiquitin (tag 1= biotin, tag 2= DNP) (human, synthetic)

UbiQ code : UbiQ-056

Batch # : B01112013-001

### Product Information

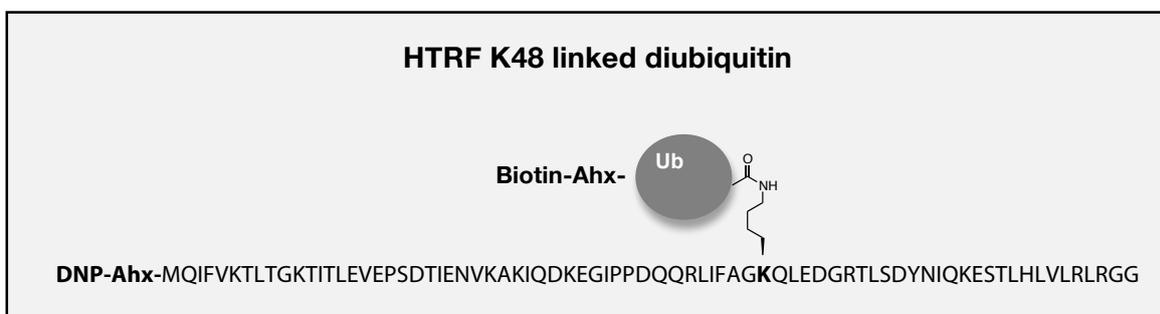
Amount : 50 ug lyophilized powder

Purity : >90% by RP-HPLC and SDS-PAGE.

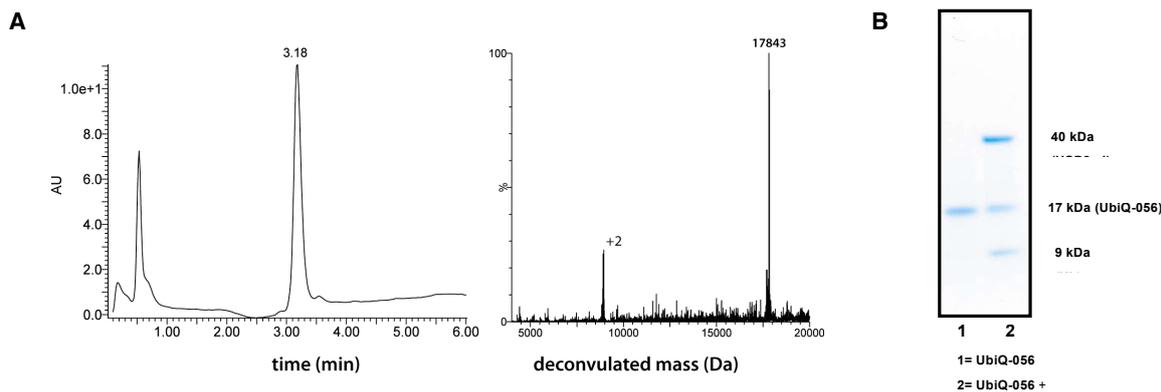
Mol. Weight : 17843 Da by MS (calc Mw 17840 Da)

Storage : powder at -20°C; solution at -80°C. Please avoid multiple freeze/thaw cycles.

**Background.** UbiQ-056 is a native K48 linked di-Ub which is equipped with a biotin on the N-terminus of the proximal ubiquitin and a dinitrophenyl group on the N-terminus of the distal ubiquitin (DNP= N-(2,4-dinitrophenyl)-6-aminohexanoic Acid). Both tags are separated from the N-terminus by an additional aminohexanoic acid (Ahx) linker. UbiQ-056 is designed to be used in an *Homogeneous Time-Resolved Fluorescence* (HTRF) assay as a substrate for proteases that cleave the isopeptide linkage between two ubiquitin molecules.<sup>1,2</sup> This product is formed by chemical ligation.<sup>3</sup>



**Important - Sample preparation.** Dissolve the powder in a minimal amount of DMSO (e.g. 0.5 mg in 25  $\mu$ L DMSO gives a stock of 1121  $\mu$ M) and add this DMSO stock slowly to milliQ or the required buffer (please note the order of addition). Buffer exchange using 3 kDa spin filters (or dialysis membrane) can be used to remove the DMSO if this is required.



**A:** LC-MS analysis. Mobile phase A= 1%  $\text{CH}_3\text{CN}$ , 0.1% formic acid in water (milliQ) and B= 1% water (milliQ) and 0.1% formic acid in  $\text{CH}_3\text{CN}$ . Phenomenex Kinetex C18, (2.1 $\times$ 50 mm), 2.6  $\mu$ M; flow rate = 0.5 mL/min, runtime = 6 min, column T = 40°C. Gradient: 5% $\rightarrow$ 95% B over 3 $\frac{1}{2}$  min. **B:** SDS-PAGE analysis and cleavage of UbiQ-056 by USP8cd. **UbiQ-056** (3 ug in 10  $\mu$ L, 18  $\mu$ M) was incubated for 30 min at 37°C with USP8cd (3.7  $\mu$ M) after which the reactions were stopped with sample buffer and boiling for 5 min. at 95°C. Proteins were separated by SDS-PAGE (12%, MES buffer) and stained with CBB G-250.

**Literature.** (1) Horton et al. *Analytical Biochemistry* **2007**, 360, 138. (2) Engels et al. *Analytical Biochemistry* **2009**, 390, 85. (3) El Oualid et al. *Angew. Chemie Int. Ed.* **2010**, 49, 10149.

**For Laboratory Research Use Only, Not For Use in Humans**

2014

Amsterdam, The Netherlands