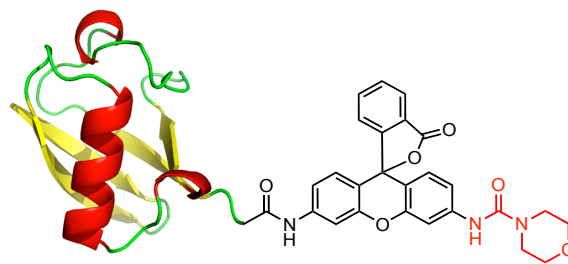


# UbiQ

targeting the ubiquitin system



## Ub-Rh110MP (human sequence, synthetic)

UbiQ code : UbiQ-126  
Batch # : B01092015-001  
Amount : 50 ug, lyophilized powder  
Purity : ≥95%  
Mol. Weight : 8.97 kDa  
Storage : upon arrival powder at -20°C, solution at -80°C. Please avoid multiple freeze/thaw cycles.

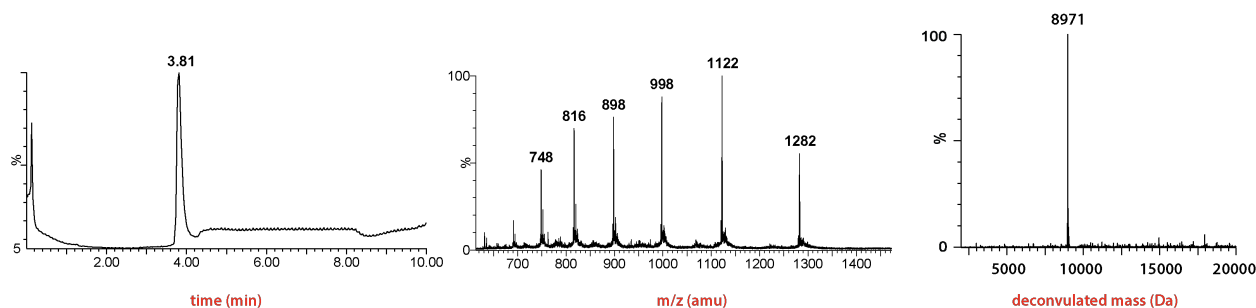
## Productsheet

**Background.** Ub-Rh110MP is a new type of quenched, fluorescent substrate for deubiquitinating enzymes (DUBs). Cleavage of the amide bond between Ub Gly76 and the Rhodamine110 moiety releases the highly fluorescent Rh110-morpholinecarbonyl (Rh110MP, exc/emi= 492/525 nm).<sup>1,2</sup> Rh110MP exhibits a higher fluorescence intensity than the classical Rh110Gly fluorophore of Ub-Rh110Gly (UbiQ-002).<sup>3</sup> UbiQ-126 is prepared by total chemical synthesis.<sup>4</sup>

- keep the excellent properties of the classic ubiquitin-Rh110 substrate (Figures 1 - 3)
- with increased fluorescence intensity after proteolytic cleavage (Figures 1C and 3).

### sequence

MQIFVKTLTGKTTILEVEPSDTIENVKAKIQDKEGIPPDQQRLLIFAGKQLEDGRTLSDYNIQKESTLHLVLRRLRGG-Rh110MP



**LC-MS analysis.** Mobile phase A= 1% CH<sub>3</sub>CN, 0.1% formic acid in milliQ and B= 1% milliQ and 0.1% formic acid in CH<sub>3</sub>CN. XBridge BEH300 C18 5µm 4.6x100mm; column T= 40°C, flow= 0.8 mL/min. Gradient: 30–60% over 6.5 min.

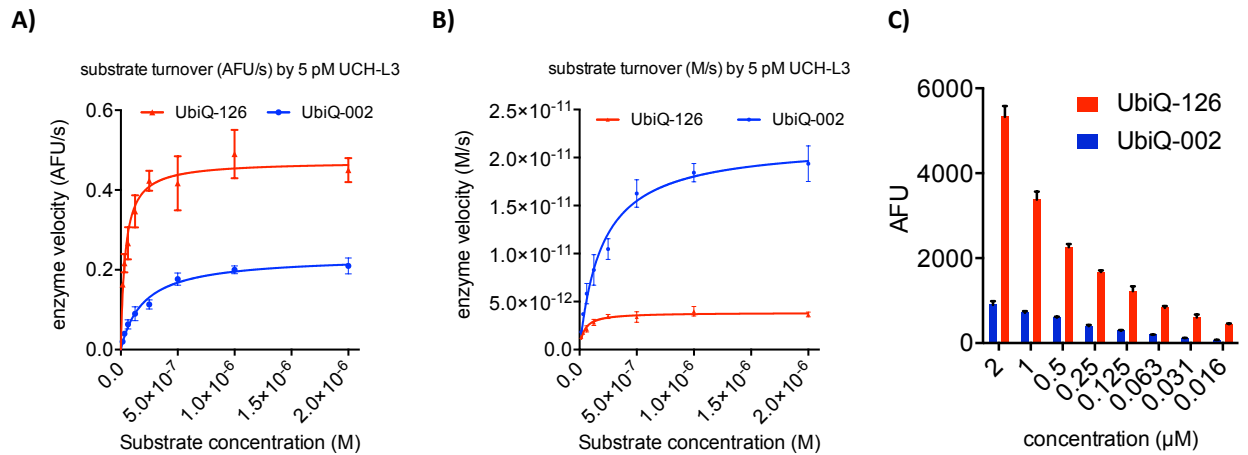
### important: sample preparation

- dissolve the powder in DMSO: DMSO stocks can range from 0.9 mg/mL (100 µM) to 40 mg/mL (4.45 mM)
- add the DMSO stock to milliQ (please note the order of addition) and mix
- buffer the aq. solution as desired (using 1M HEPES or 1M Tris for example)
- a final assay stock of 100 nM will contain 0.1 vol% DMSO when prepared from a 100 µM DMSO stock

**Literature.** (1) Lavis et al. *ACS Chem Biol* **2006**, *1*, 252. (2) Terentyeva et al. *Bioconj Chem* **2011**, *22*, 1932. (3) Hassiepen et al. *Analytical Biochem* **2007**, *371*, 201. (4) El Oualid et al. *Angew Chem Int Ed* **2010**, *49*, 10149.

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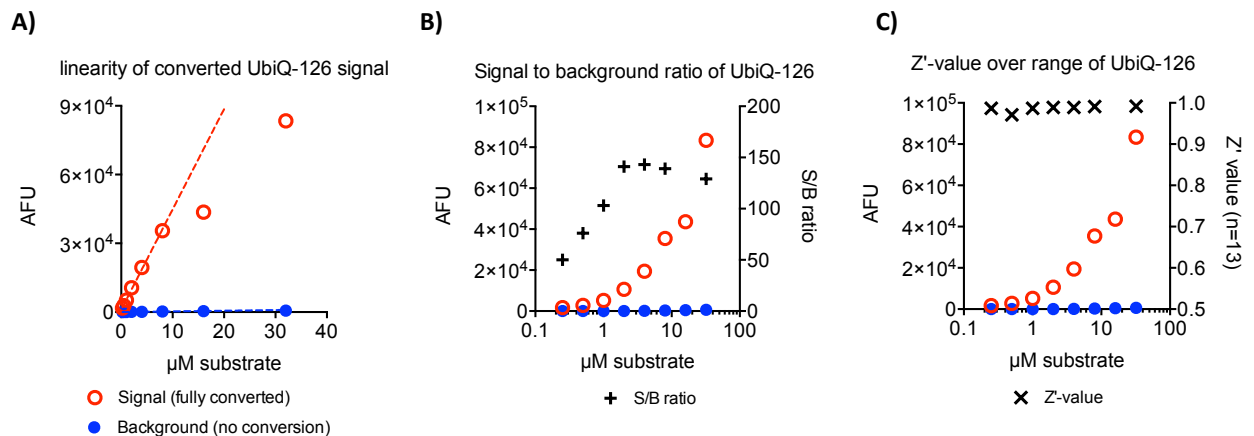


**Figure 1. Michaelis-Menten kinetics of UbiQ-126 (Ub-Rh110MP) and UbiQ-002 (Ub-Rh110Gly), turned over by 5 pM UCH-L3.** Enzyme kinetics were determined in 384 well format (30 uL per well) on a *BMG Clariostar plate reader* measuring fluorescence intensity at  $\lambda_{exc}$  487 ± 14 nm;  $\lambda_{emi}$  535 ± 30 nm; 40 flashes per well.

A) enzyme velocity represented as AFU/s versus substrate concentration. AFU: arbitrary fluorescence units.

B) enzyme velocity represented as M/s versus substrate concentration.

C) fluorescence intensities determined at 30 min turnover by 5 pM UCH-L3, error bars are SD (n=3).



**Figure 2. Fluorescence signal versus background of UbiQ-126 (Ub-Rh110MP).** Fluorescence intensities were measured of various concentrations of **UbiQ-126** (background) and fully converted **UbiQ-126** by 1 μM USP7 (signal).

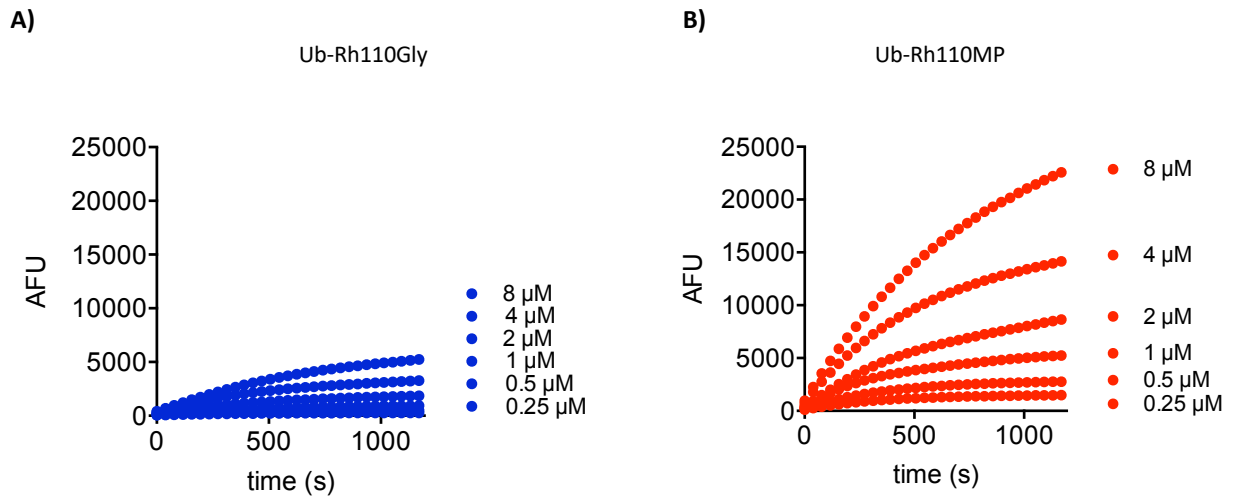
A) the fluorescence signal of processed **UbiQ-126** is linear up to 8 μM.

B) signal-to-background ratios over a concentration range of **UbiQ-126**.

C) Z'-values over a concentration range of **UbiQ-126**, determined over 13 replicates. Fluorescence intensities were measured in 384 well format on a *BMG Pherastar plate reader* at  $\lambda_{exc}$  485 ± 16 nm;  $\lambda_{emi}$  520 ± 10 nm.

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**Figure 3. Progress curves of the conversion of UbiQ-002 (Ub-Rh110Gly) and UbiQ-126 (Ub-Rh110MP) by 1 nM USP7.** Fluorescence intensities were measured in 384 well format on a *BMG Pherastar plate reader* at  $\lambda_{\text{exc}}$  485  $\pm$  16 nm;  $\lambda_{\text{emi}}$  520  $\pm$  10 nm. AFU: arbitrary fluorescence units.  
A) conversion of **Ub-Rh110Gly** (blue dots) at the indicated concentrations.  
B) conversion of **Ub-Rh110MP** (red dots) at the indicated concentrations.